



## Data in Brief

## De novo transcriptome assembly of loggerhead sea turtle nesting of the Colombian Caribbean

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## ARTICLE INFO

## ABSTRACT

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Loggerhead sea turtle *Caretta caretta* is widely distributed in the oceans of tropical and subtropical latitude. This turtle is an endangered species due to anthropic and natural factors that have decreased their population levels. In this study, RNA sequencing and de-novo assembly of genes expressed in blood were performed. The raw FASTQ files have been deposited on NCBI's SRA database with accession number SRX2629512. A total of 5.4 Gb raw sequence data were obtained, corresponding to 48,257,019 raw reads. Trinity pipeline was used to perform a de-novo assembly, we were able to identify 64,930 transcripts for female loggerhead turtle transcriptome with an N50 of 1131 bp. The obtained transcriptome data will be useful for further studies of the physiology, biochemistry and evolution in this species.

## Specifications

Organism/cell line/tissue	<i>Caretta caretta</i> /blood tissue
Sex	Female
Sequencer or array type	Illumina Hiseq2000
Data format	Raw data FASTQ file
Experimental factors	De novo transcriptome assembly of <i>Caretta caretta</i>
Experimental features	A blood sample of a living specimen of the sea turtle <i>Caretta caretta</i> was collected for total RNA isolations. Prepared a paired-end library, sequenced by the Hiseq 2000 system. The obtained data was subjected to de novo assembly.
Consent	N/A
Sample source location	Islas del Rosario, Bolivar, Colombia

## 1. Direct link to deposited data

<https://www.ncbi.nlm.nih.gov/sra/SRX2629512>

## 2. Introduction

The loggerhead turtle *Caretta caretta* [19], is distributed around the oceans of the world in tropical and subtropical latitudes [1]. The main nesting beaches are found in the Florida Peninsula in North America [2], Brazil, Japan and Greece and on the Mediterranean Sea, Oman in the Arabian Sea, and on the Madagascar Island [3–5]. It is an important member of complex ecological marine and coastal systems [6]. It contributes to the resilience of marine environments maintaining the balance in ecosystems and food chains they occupy, through the control of mollusks, crustaceans and other invertebrate marine populations [7]. Its main contribution is to transfer large amounts of biomass to abyssal zones [8]. In the Colombian Caribbean, the presence of this species has been recognized, and nesting beaches in the departments of Magdalena, Guajira, Bolivar and San Andres have been identified [9]. *Caretta caretta* is protected by national laws and international agreements that hopefully will mitigate the decrease of nesting females caused by anthropic causes [10,11]. However, Colombia has the second highest level of *Caretta caretta* capturing per year in the world, with an approximate

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number/year of 600 turtles [12]. In 1986, the International Union for Conservation of Nature [IUCN] reported the *Caretta caretta* turtle in its red list and denoted as vulnerable; in 1996, it entered to the endangered (EN) category; currently it can be found in the category of vulnerable A2b (11). The population decline of the *Caretta caretta* turtle is attributed to factors such as contamination, modification of the habitat, introduction of predators, by-catch and excessive clandestine fishing [13]. In addition, pathologies such as pneumonia, hepatitis, meningitis, eye conditions and other conditions associated with exposure to hydrocarbons and organochlorine pesticides have been described, which cause the death of loggerhead turtles [7]. Other pathologies such as malformations, depigmentation and embryonic mortality have not yet been studied, but could be related to genetic factors [14].

The metabolic adaptations and physiological mechanisms underlying their ability to move long distances, prolonged anoxia times and proper water maintenance have been the aim of intense interest for many years [15]. Loggerhead turtles generally exhibit contrary behavior to divers who breathe air, they rest in the background, regularly for long periods during the demersal stages of their life cycle. They have high anaerobic capacity and tolerance to anoxia. Given that lungs collapse during deep dives, their storage capacity is expendable, species increase their blood volume and concentrations of respiratory pigments [15]. Additionally, the tolerance to anoxia is developed as they increase in size, which defines their behavior in the water column.

In this study, we performed de novo transcriptome assembly for a blood sample from *Caretta caretta* sea turtle from young female individual by next-generation sequencing. This transcriptome data will be useful for further studies of the physiology, biochemistry and evolution.

### 3. Experimental design, materials and methods

#### 3.1. Animal materials

Blood tissue sample from a *Caretta caretta* individual was obtained from the CEINER Oceanarium in San Martin de Pajares Island, Cartagena. The blood was obtained from the dorsal cervical breasts in accordance with [20] methodology. The sample was placed in sterilized tubes with Tris-EDTA buffer 0.1 M (GreinerBio-one®, Kremsmünster, Austria) solution and was transported at 4 °C to the Molecular Biology lab of the Universidad Jorge Tadeo Lozano, Bogota campus. The sample was collected following the ethical standards established by the legislation and the study obtained permission from the Ministry of the Environment for the development of the Biodiversity research (No 24 of June 22, 2012) and the Genetic Resources Access contract (No 64 of April 2013).

A blood sample of a *Caretta caretta* sea turtle was used for total RNA extraction using RNeasy Mini Kit (Quiagen, Hilden, Germany). For mRNA library preparation, we use a TruSeq RNA Library Prep Kit v2 according to manufacturer's instructions (Illumina, San Diego, U.S.A.). The poly-A containing mRNAs were isolated using poly-T oligo-attached magnetic beads. The first strand cDNA followed by a second strand cDNA was synthesized from purified mRNAs. End repair was performed followed by adenylation of 3 ends. Adapters were ligated and PCR was done to selectively enrich DNA fragments with adapters and to amplify the amount of DNA in the library, respectively. The quality control of generated libraries was done using the 2100 bio-analyzer (Agilent, Santa Clara, U.S.A.). RIN values (RNA integrity number) of 7.5 were obtained. The library was paired-end sequenced by Macrogen Co. (Seoul, South Korea) using Hiseq 2000 Platform. The quality of cleaned raw reads was verified with the fastQC program (<http://www.bioinformatics.bbr.ac.uk/projects/fastqc/>).

#### 3.2. De novo transcriptome assembly

The quality of sequencing reads was performed by means of FastQC [16]. Read trimming on quality (Q50) and sequencing adaptors

**Table 1**  
Statistics of *Caretta caretta* transcriptome assembly.

No of raw data bases	4,873,958,919
No of reads of pairs	48,257,019
No of assembled transcripts	64,930
Assembly GC percent	45.8
Contig N50	1731
Contig minimum	165
Contig maximum	11,046
Average contig	731

removal was run with Trimmomatic [17]. We obtained a total of 5,4 Gb of raw data corresponding to 4,873,958,919 bp.

De novo transcriptome assembly was performed using Trinity [18], this program, was executed using default parameters for the assembly of paired end reads. Mapping and abundance estimation was performed by means of Bowtie [21] using the constructed transcriptome as a reference. Transcriptome sequencing and read processing are summarized in Table 1. We obtained a total of 64,930 assembled transcripts with a N50 = 1731 bp, average length of 731 bp.

In conclusion, hereby we present the first sequencing effort and the novo assembly of the transcriptome of the loggerhead sea turtle *Caretta caretta*. This transcriptome data will be useful for further studies of the evolution, phylogenomics, physiology, biochemistry and gene regulation of anoxia tolerance.

#### Conflict of interest

The authors declare that they have no competing interests.

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